Bacteriological Profile and Antimicrobial Susceptibility Pattern among Isolates Obtained From Body Fluids

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ABSTRACT

Background: Infections of the sterile body sites, when occurs typically have greater clinical urgency and these infections could be life threatening. For the appropriate management of patient, early detection and identification of organism is crucial. Hence, the present study was designed to evaluate the prevalence, organism profile and antimicrobial susceptibility pattern of isolates obtained from infection of body fluids.

Methods: Laboratory and clinical records of all the sterile body fluid except blood and cerebrospinal fluid submitted to the Department of Microbiology, B. P. Koirala Institute of Health Sciences during the period of 5 years (January 2012 to December 2016) were analyzed.

Results: Among 1835 samples, 196 fluids samples showed growth of organisms with an isolation rate of 10.68%. *Escherichia coli (E. coli)* was the most common organism followed by *Staphylococcus aureus (S. aureus)* and *Acinetobacter calcoaceticus-baumannii complex*. Antimicrobial susceptibility testing showed variable degree of resistance. Thirty percentage of organisms were multi-drug resistant, 10% were extensively-drug resistant, none was pan-drug resistant, 35% of *E. coli* and *K. pneumoniae* were extended spectrum β -lactamase producer and 30% of *S. aureus* were methicillin-resistant *S. aureus*. The study showed increasing trends of multi-drug resistant, extensively-drug resistant, methicillin-resistant *S. aureus* and extended spectrum β -lactamase over the years.

Conclusions: The study showed infection rate of 10% among normally sterile body fluids. *E.coli, S. aureus* and *Acinetobacter calcoaceticus-baumannii complex* are the common organisms. The infections are associated with multi drug resistant organisms. Routine surveillance of multi-drug resistant in infection of body fluids is necessary to guide treatment.

Keywords: Extensively-drug resistant; methicillin-resistant S. aureus; multi-drug resistant; sterile body fluid.

INTRODUCTION

Body fluids like pleural, ascitic, peritoneal, synovial and pericardial fluid are usually sterile. Infections of the sterile body sites, when occurs typically have greater clinical urgency and these infections could be life threatening.^{1,2} For the appropriate management of patient, early detection and identification of organism with the results of antimicrobial susceptibility testing is crucial. Positive cultures are usually low because of less number of pathogens and prior administration of empirical antibiotics in these patients. Moreover, the emergence of antimicrobial resistance especially multidrug resistant (MDR), extensively-drug resistant (XDR), pandrug resistant (PDR) organisms, methicillin-resistant *Staphylococcus aureus* (MRSA)and extended-spectrum B-lactamase (ESBL)producers has hindered the clinical management of the patient.^{3,4}

Regular monitoring of bacterial susceptibility pattern a particular area is necessary for empirical treatment of infection as soon as possible, which helps in reduction of morbidity and mortality.⁵ Hence, the current study was designed to evaluate the prevalence, organism profile and antimicrobial susceptibility pattern of isolates obtained from infection of body fluids.

METHODS

This is a hospital based retrospective study conducted in the Department of Microbiology, BPKIHS. Ethical clearance was obtained from Institutional review board, BPKIHS. Laboratory and clinical records of all patients admitted during the period of 5 years (January 2012

Correspondence: Lok Bahadur Shrestha, Department of Microbiology & Infectious Diseases, B. P. Koirala Institute of Health Sciences, Dharan, Sunsari, Nepal. Email:lok.shrestha@bpkihs.edu, Phone: +9779842295909. to December 2016) whose body fluid(except blood and cerebrospinal fluid) yielded positive growth was traced and analyzed. An attempt was made to establish the clinical significance of the isolates with clinical corelations and records of repeat culture and sensitivity.

Body fluid specimens when received in microbiology laboratory were subjected to gram staining and culture. The specimen is inoculated onto Blood agar, MacConkey agar and Chocolate agar and incubated aerobically at 35 °C for 18-24 hours. After incubation, the plates were observed for bacterial growth. Any bacterial colony was identified by using gram staining and biochemical tests following standard microbiological guidelines.⁶After identification, antimicrobial susceptibility testing was performed by modified Kirby Bauer disc diffusion method following clinical and laboratory standards institute guidelines.⁷

MDR is defined as acquired non-susceptibility to at least one agent in three or more antimicrobial categories and XDR is defined as non-susceptibility to at least one agent in all but two or fewer antimicrobial categories (i.e. bacterial isolates remain susceptible to only one or two categories) and PDR is defined as non-susceptibility to all agents in all antimicrobial categories.⁸

RESULTS

During the study period, samples from 1835 patients were submitted for culture and sensitivity. Among the patients, 54 % were male while 46% were female. Age-wise distribution of the patient showed 12% were children, 52% were adults and 36% were elderly. Among them, samples from 196 patients (10.68%) showed growth.Total number of samples and growth rate has been further elicited in Table 1.

Table 1. Number of samples and growth rate.							
Samples	Total no of	Culture	Growth				
	samples	positive	percentage				
Pleural fluid	1172	137	11.68				
Ascitic fluid	517	42	8.12				
Synovial fluid	125	15	12				
Pericardial fluid	21	2	9.5				
Total	1835	196	10.68				

Among 137 bacterial isolates obtained from pleural fluid, *Escherichia coli* (n=34, 25%) was most common followed by *Acinetobacter calcoaceticus-baumannii complex* (*ACB complex*)(n=25, 18%).Similarly, *E. coli*(n=12, 28%) was also the most common organism isolated from ascitic fluid followed by *Staphylococcus aureus* (n=7, 16%). Whereas, *S. aureus* (n=7, 46%)was the most common isolate obtained from synovial fluid followed by *E. coli* (n=4, 26%). *S. aureus*(n=2) was the only bacteria isolated from pericardial fluid (Table 2).

Antimicrobial susceptibility pattern of the isolates has been tabulated in Table 3. Eighty percentage of *E. coli* were resistant to ampicillin while only 5% resistance was seen towards meropenem. Among gram-positive bacteria, 90% of *S. aureus* were resistant to penicillin while all were susceptible to vancomycin and linezolid.

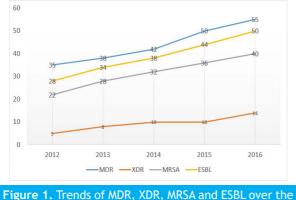
Table 2.Organisms isolated from different clinical samples.								
Organism	Pleural fluid	Ascitic fluid	Synovial fluid	Pericardial fluid	Total			
Escherichia coli	34	12	4	-	50			
Klebsiella pneumoniae	14	3	-	-	17			
Citrobacter freundii	5	3	-	-	8			
Enterobacter aerogenes	13	4	-	-	17			
Pseudomonas aeruginosa	10	4	-	-	14			
ACB complex	25	5	1	-	31			
Staphylococcus aureus	24	7	7	2	40			
Enterococcus faecalis	10	4	1	-	15			
Streptococcus pneumoniae	2	-	2	-	4			
Total	137	42	15	2	196			

Note: ACB complex: Acinetobacter calcoaceticus-baumannii complex

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Table 3. Antimicrobial resistance pattern of the isolates (%).												
Organism	Amikacin	Ampicillin	Ceftriaxone	Cefoxitin	Ofloxacin	Gentamicin	Co-trimoxazole	ΡΙΤ	Meropenem	Penicillin	Vancomycin	Linezolid
Escherichia coli	20	80	50	-	60	30	50	20	5	-	-	-
Klebsiella pneumoniae	40	-	70	-	65	45	-	30	10	-	-	-
Citrobacter freundii	10	60	65	-	70	75	-	20	20	-	-	-
Enterobacter aerogenes	15	70	65	-	65	72	-	25	20	-	-	-
Pseudomonas aeruginosa	25	-	80	-	80	55	-	40	20	-	-	-
ACB complex	40	-	75	-	60	50	-	40	20	-	-	-
Staphylococcus aureus	20	-	50	30	55	30	70	-	-	90	0	0
Enterococcus faecalis	-	-	-	-	60	35	-	-	-	70	0	0
Streptococcus pneumoniae	0	-	0	-	0	0	0	-	-	0	0	0

Note: PIT: Piparacillin-Tazobactam, ACB complex: Acinetobacter calcoaceticus-baumanniicomplex.



years (Percentage).

Note: MDR: multi-drug resistant, XDR: extensively-drug resistant, MRSA: methicillin- resistant *Staphylococcus aureus*, ESBL: extended spectrumß-lactamase

Out of 196 isolates, thirty percentof organisms were MDR, 10% were XDR, 35% of *E. coli* and *K. pneumoniae* were ESBL producer and 30% of *S. aureus* were MRSA. None of the isolates was PDR. The study showed increasing trends of MDR, MRSA and ESBL over the years (Figure 1).

DISCUSSION

Infections of the sterile body sites, when occurs is a medical emergency and these infections have high morbidity and mortality. Despite its importance, very few studies have been conducted regarding the prevalence and antimicrobial susceptibility pattern of these infections. We analyzed the laboratory records of 1835 body fluid samples submitted for culture and sensitivity that yielded 10.68% positive growth (n=196).

A study conducted by Sujatha R et al.² in Kanpur, India showed that 31% of body fluids yielded growth. Similar type of growth rate was obtained by Sharma et al (30%),⁵ and Deb et al (21%).⁹ The difference might be attributed to the long study period and large sample size in our study.

Among total samples, 64% (n=1172) were pleural fluid, 28% (n=517) ascitic fluid, 7% (n=125) synovial fluid and 1% (n=21) were pericardial fluid. Among 196 positive growth, *E. coli* (n=50, 25%) was the most common organism isolated followed by *S. aureus* (n=40, 20%) and *ACB complex* (n=31,16%). Similar results were obtained by Sharma R et al⁵ and Sheikhbahaei et al.¹⁰A study conducted by Sujatha R et al. in India also isolated *E. coli* as the most common organism causing infection of body fluids.In contrast to the finding of our study, Deb A et al. isolated *Pseudomonas aeruginosa* as the most common organism causing body fluid infection followed by *A. anitratus.⁹E. coli* is the most common bacteria causing body fluid infections.¹¹

Antimicrobial susceptibility testing showed variable degree of resistance among organisms. Eighty percent of *E. coli*were resistant to ampicillin, 60% to ofloxacin and 50% to ceftriaxone. However, only 5% of *E. coli* were resistant to meropenem. Among Gram-positive bacteria, all *S. aureus* were susceptible to vancomycin and linezolid, while 90% of them were resistant to penicillin and 70% to co-trimoxazole. *S. pneumoniae* were susceptible to all the antimicrobial agents tested. Similar pattern of antimicrobial susceptibility was reported in various studies (Sujatha et al.², Sharma

et al.⁵). This high resistance level may be due to the inappropriate use of commonly prescribed antibiotics.¹²

Fourty percentage of organisms were MDR, 10% were XDR, 37% of E. coli and K. pneumoniae were ESBL producer and 30% of S. aureus were MRSA. A study conducted by Basak S et al.¹³ observed MDR in 37% isolates, XDR in 13% of the isolates, 31% MRSA which is guite similar to the finding of our study. They did not isolate any PDR organisms, which also agrees to our finding. However, they detected ESBL production in 18% of the Gramnegativebacilli, which is lesser than our finding (37%). A study conducted by Shrestha A et al.¹⁴ in Chitwan, Nepal observed 79% MDR and 36% ESBL producing GNB. Our study also showed the increasing trend of MDR, XDR, ESBL and MRSA over the years. Similar results have been reported by studies done worldwide.^{4,15} There has been a worldwide increase in emergence of drug resistant organisms in recent years.¹⁵

MDR bacteria has been well recognized as one of the most important public health problems in current scenario. Treatment outcomes in patients infected with MDR bacteria tend to be worse as compared to those infected with susceptible organisms.³ MRSA is probably the best example of a prevalent and important MDR bacterium that has successfully transitioned from an almost exclusively nosocomial setting to being widespread in the community. Several researches have concluded that MRSA is increasing as a cause of community-acquired infections.¹⁶⁻¹⁸Worldwide, the prevalence of MRSA range from 30% to 90% depending upon the type of infections.^{3,19} A study conducted at the National Public health laboratory (NPHL), Kathmandu, Nepal reported that 31.57% of E. coli were confirmed as Extended Spectrum B-lactamase producers.²⁰ The Prevalence of ESBL producer worldwide range from 12 to 80%. The epidemiology of ESBL-producing bacteria is becoming more complex with increasingly blurred boundaries between hospitals and the community.²¹

CONCLUSIONS

The study showed infection rate of 10% among normally sterile body fluids. *E. coli*, *S. aureus* and *ACB complex* are the common organisms. There is an increasing trend of antimicrobial resistance. Routine surveillance of MRSA, ESBL, and multi-drug resistant organisms is essential in proper management of body fluid infections.

REFERENCES

1. Hughes JG, Vetter EA, Patel R, Schleck CD, Harmsen S, Turgeant LT, et al. Culture with BACTEC Peds

Plus/F bottle compared with conventional methods for detection of bacteria in synovial fluid. J Clin Microbiol. 2001;39(12):4468-71.[PubMed]

- Sujatha R, Nidhi P, Arunagiri D, Narendran D. Bacteriological profile and antibiotic sensitivity pattern from various body fluids of patients attending Rama medical college hospital, Kanpur. Int J Advances in Case Reports. 2015;2(3):119-24.
- van Duin D, Paterson DL. Multidrug-Resistant Bacteria in the Community: Trends and Lessons Learned. Infect Dis Clin North Am. 2016;30(2):377-90.[PubMed]
- McDanel J, Schweizer M, Crabb V, Nelson R, Samore M, Khader K, et al. Incidence of Extended-Spectrum beta-Lactamase (ESBL)-Producing Escherichia coli and Klebsiella Infections in the United States: A Systematic Literature Review. Infect Control Hosp Epidemiol. 2017;38(10):1209-15.[PubMed]
- Sharma R, Anuradha., Nandini D. Bacteriological Profile and Antimicrobial Sensitivity Pattern in Sterile Body Fluids from a Tertiary Care Hospital. J Appl Microbiol Biochem. 2017;1(1).
- Procop GW, Church GL, Hall GS, Janda WM, Koneman EW. Koneman's Color Atlas and Textbook of Diagnostic Microbiology. PA: Wolters Kluwer; 2016.
- Clinical and Laboratory Standards Institute. CLSI document M100S. Performance Standards for Antimicrobial Susceptibility Testing. 26th ed. Wayne, PA. 2016.
- Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. Clin Microbiol Infect. 2012;18(3):268-81.[PubMed]
- Deb A, Mudshingkar S, Dohe V, Bharadwaj R. Bacteriology of Body Fluids with an Evaluation of Enrichment Technique to Increase Culture Positivity. J Evolution Med and Dental Sci. 2014;3(72):15230-8.[FullText]
- Sheikhbahaei S, Abdollahi A, Hafezi-Nejad N, Zare E. Patterns of antimicrobial resistance in the causative organisms of spontaneous bacterial peritonitis: a single centre, six-year experience of 1981 samples. Int J Hepatol. 2014;2014:917856.[PubMed]
- Johnson JR. Virulence factors in Escherichia coli urinary tract infection. Clin Microbiol Rev. 1991;4(1):80-128.
 [PubMed]
- 12. Kardas P, Devine S, Golembesky A, Roberts C. A systematic review and meta-analysis of misuse of antibiotic therapies in the community. Int J Antimicrob Agents.

2005;26(2):106-13.[PubMed]

- Basak S, Singh P, Rajurkar M. Multidrug Resistant and Extensively Drug Resistant Bacteria: A Study. J Pathog. 2016;2016:4065603.[PubMed]
- 14. Shrestha A, Bajracharya AM, Subedi H, Turha RS, Kafle S, Sharma S, et al. Multi-drug resistance and extended spectrum beta lactamase producing Gram negative bacteria from chicken meat in Bharatpur Metropolitan, Nepal. BMC Res Notes. 2017;10(1):574.[PubMed]
- Exner M, Bhattacharya S, Christiansen B, Gebel J, Goroncy-Bermes P, Hartemann P, et al. Antibiotic resistance: What is so special about multidrug-resistant Gram-negative bacteria? GMS Hyg Infect Control. 2017;12:Doc05. [PubMed]
- Bukharie HA. Increasing threat of community-acquired methicillin-resistant Staphylococcus aureus. Am J Med Sci. 2010;340(5):378-81.[PubMed]
- Mera RM, Suaya JA, Amrine-Madsen H, Hogea CS, Miller LA, Lu EP, et al. Increasing role of Staphylococcus aureus and community-acquired methicillin-resistant Staphylococcus aureus infections in the United States: a 10-year trend of replacement and expansion. Microb Drug Resist. 2011;17(2):321-8. [PubMed]

- Shrestha LB, Baral R, Poudel P, Khanal B. Clinical, etiological and antimicrobial susceptibility profile of pediatric urinary tract infections in a tertiary care hospital of Nepal. BMC Pediatr. 2019;19(1):36.[PubMed]
- DeLeo FR, Otto M, Kreiswirth BN, Chambers HF. Community-associated meticillin-resistant Staphylococcus aureus. Lancet. 2010;375(9725):1557-68.[PubMed]
- Thakur S, Pokhrel N, Sharma M. Prevalence of multidrug resistant enterobacteriaceae and extended spectrum β lactamase producing Escherichia coli in urinary tract infection. RJPBCS. 2013;4(2):1615-24.[DOI]
- Shaikh S, Fatima J, Shakil S, Rizvi SM, Kamal MA. Antibiotic resistance and extended spectrum betalactamases: Types, epidemiology and treatment. Saudi J Biol Sci. 2015;22(1):90-101.[PubMed]